

**Genetic Variation of the Endangered  
*Ladigesocypris ghigii* Populations  
Revealed by the RAPD-PCR  
Technique and the RFLP Analysis of  
PCR Amplified mtDNA Segments:  
Implementation to Species  
Conservation**

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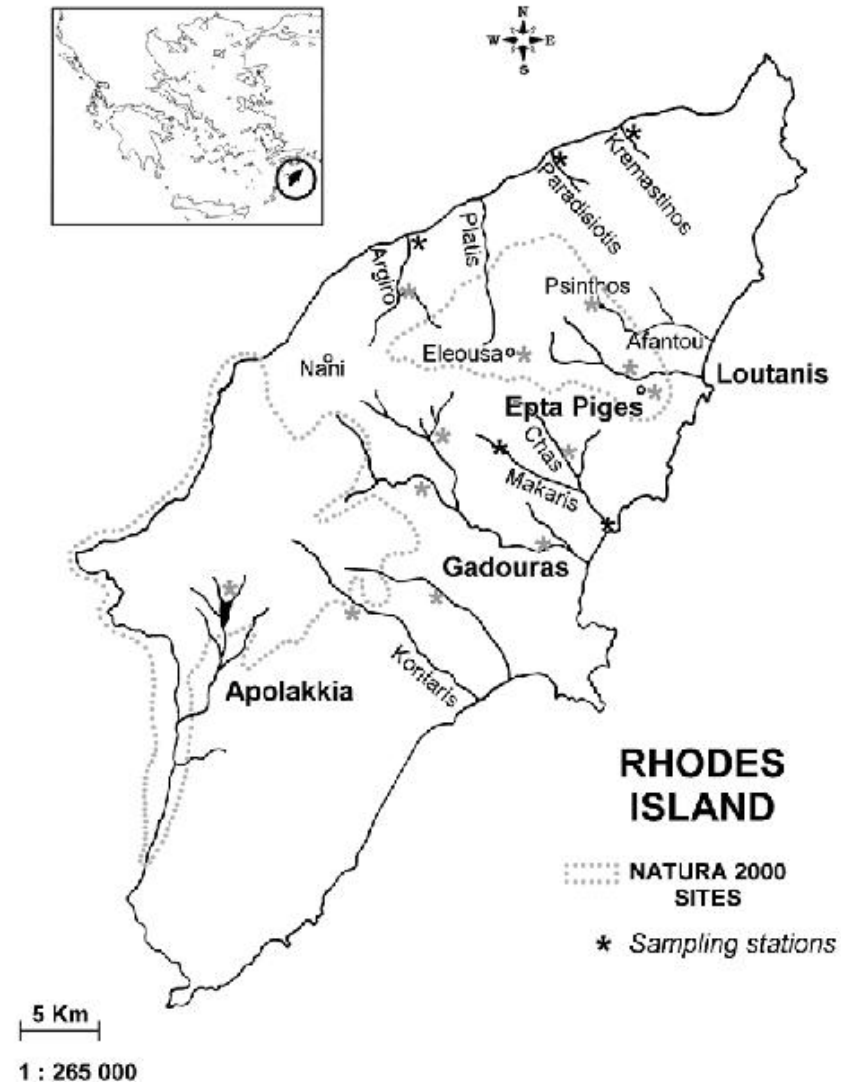
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**A freshwater fish, commonly known as "gizani" endemic to Rhodes island**

**A small-bodied cyprinid that inhabits the streams, springs and some small water reservoirs**



✓ During summers, the majority of the ravines of the island are drying, almost throughout their entire course.

✓ Small groups of fish survive in water pockets along the streams and/or near the streams' sources



Loutanis stream, June 2000



Young fishes in Loutanis,  
June 2000

Gradual decrease of water level in  
Apolakkia lake



A. April 99



B. June 00



C. September 00



D. May 01

## Therefore:

- Gizani is listed as an endangered species of top priority for conservation in ANNEX II of the Habitats Directive (92/43/EEC) of the E.U., concerning the protection of natural habitats and wild fauna and flora

## And

- A LIFE-Nature project was undertaken (B4 3200/98/445) aiming at studying the species and implementing measures for its conservation

# **In the course of this project a biological survey was undertaken to:**

- ✓ **Create an updated record of existing gizani populations (number and distribution).**
- ✓ **Define the state of the populations and the environmental threats.**
- ✓ **Determine genetic diversity and conservation units.**
- ✓ **Define bottlenecks to recruitment at all life stages.**
- ✓ **Identify options for the conservation of the species.**



It was considered essential to obtain **GENETIC INFORMATION** about the gizani populations, to define the conservation units for this species

Nuclear DNA analysis



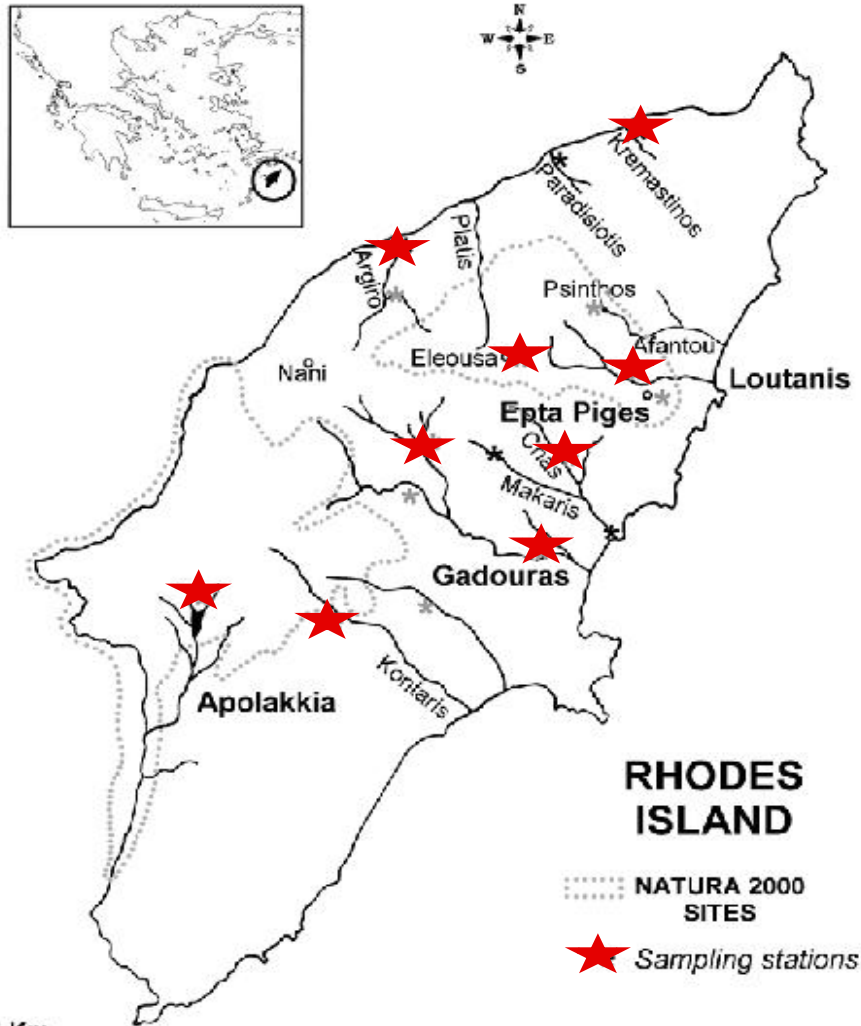
RAPD method

Mitochondrial DNA analysis

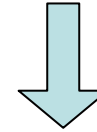


RFLP method

# The aquatic systems under study and the sampling stations

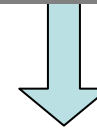


**RAPD**



**Sixteen decamer primers**

**mtDNA**



**PCR-amplified regions  
(ND-5/6, COI, 12S-16S  
rRNA) screened with 20  
restriction endonucleases,  
4-, 5-, 6-base cutters**

5 Km

1 : 265 000

## RAPD analysis

**130** bands analysed for all the 16 random primers

**6 (4.62%)** were polymorphic, observed in different frequencies in the nine populations

**112** were monomorphic, constantly present in all individuals

**12** were diagnostic (i.e. present in all individuals of one population and absent from all individuals of another population and vice-versa)



# RAPD profiles after amplification with two different primers

**A**

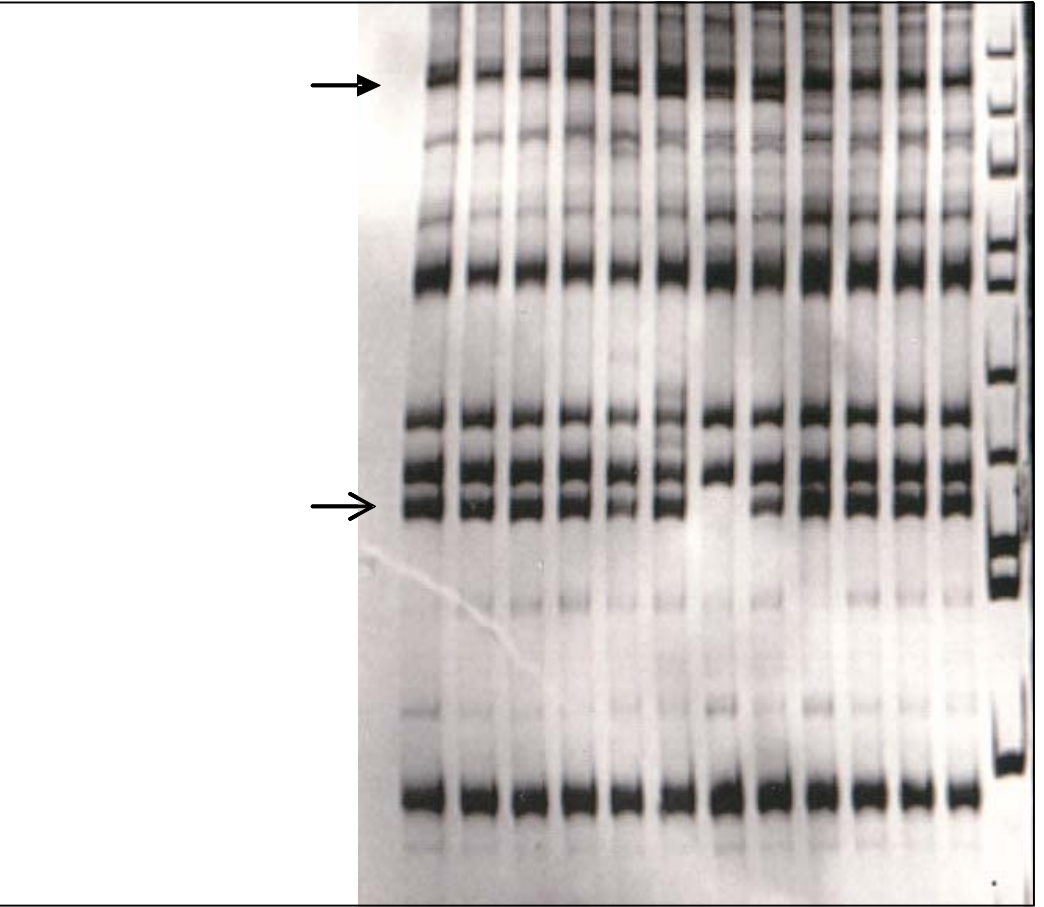
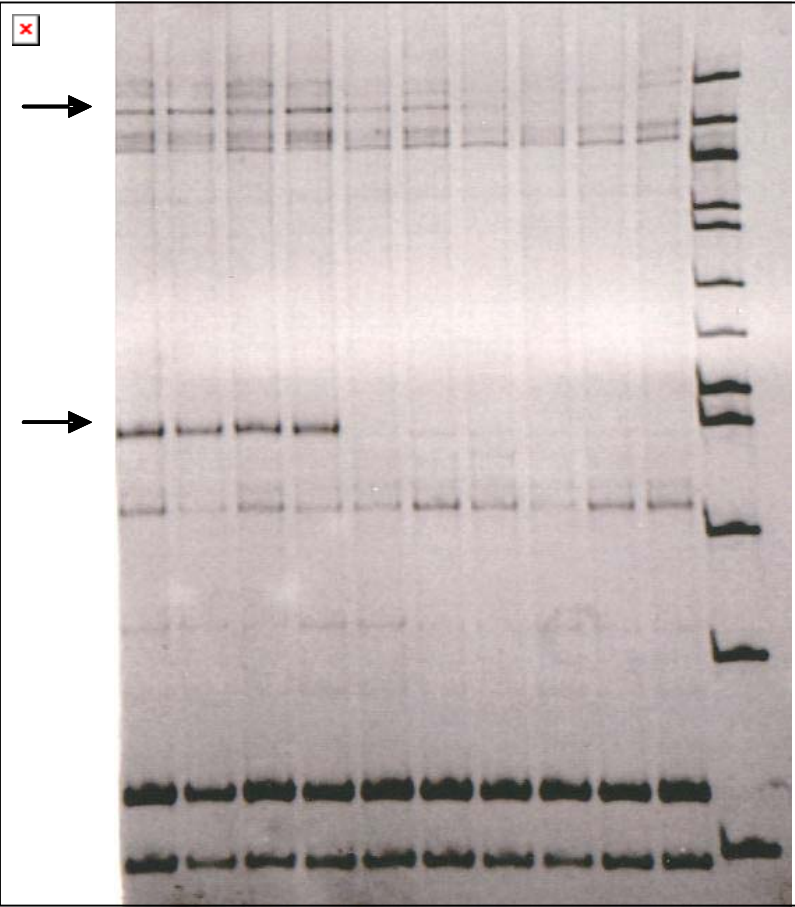
Argiro  
Gadouras  
Loutanis  
Apolakkia  
Ladder

1 2 3 4 5 6 7 8 9 10 M

**B**

Argiro  
Gadouras  
Loutanis  
Apolakkia  
Ladder

1 2 3 4 5 6 7 8 9 10 11 12 M





# mtDNA-RFLP analysis

ND-5/6 → 2.50 kb

COI → 1.30 kb

12S-16S rRNA → 2.05 kb

40% of the  
mitochondrial  
genome

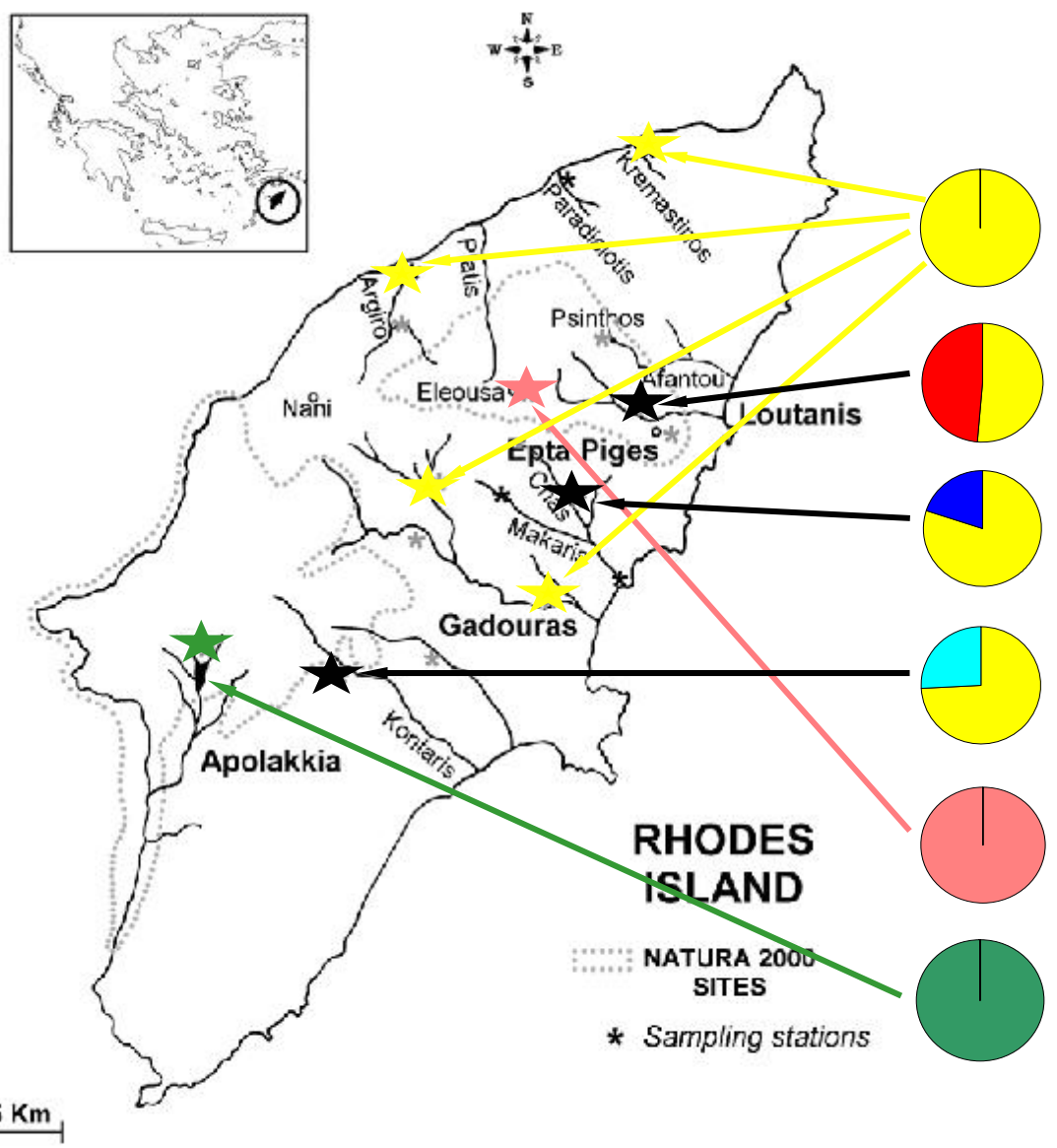
169 restriction sites → 739bp surveyed

# mtDNA-RFLP analysis

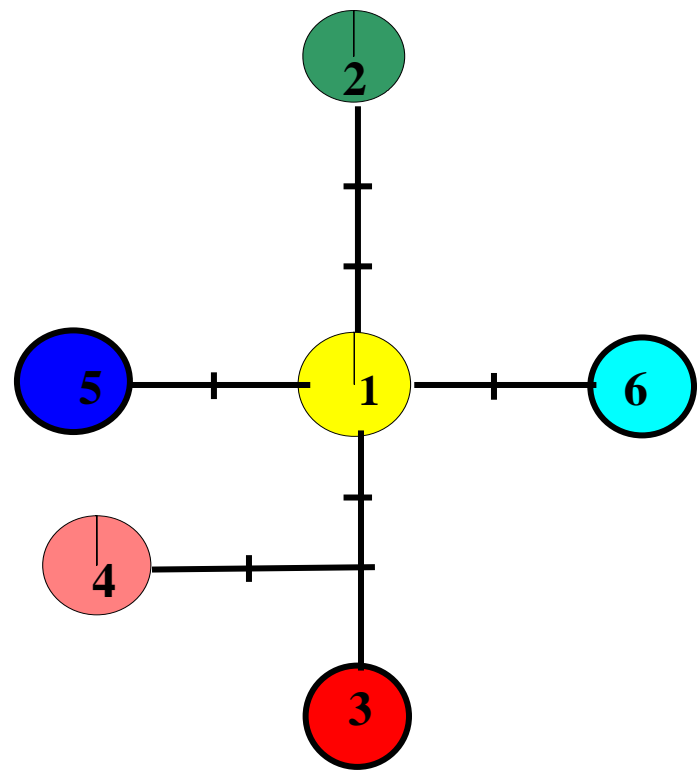
Six haplotypes with very shallow pairwise sequence divergence ( $\times 10^2$ ) (average of 0.500)

| Haplotypes | 1    | 2    | 3    | 4    | 5    | 6 |
|------------|------|------|------|------|------|---|
| 1. Type 1  | -    |      |      |      |      |   |
| 2. Type 2  | 0.41 | -    |      |      |      |   |
| 3. Type 3  | 0.41 | 0.83 | -    |      |      |   |
| 4. Type 4  | 0.41 | 0.82 | 0.69 | -    |      |   |
| 5. Type 5  | 0.21 | 0.63 | 0.63 | 0.63 | -    |   |
| 6. Type 6  | 0.21 | 0.62 | 0.62 | 0.62 | 0.42 | - |

# Pie diagrams indicating mtDNA haplotype frequencies at each site

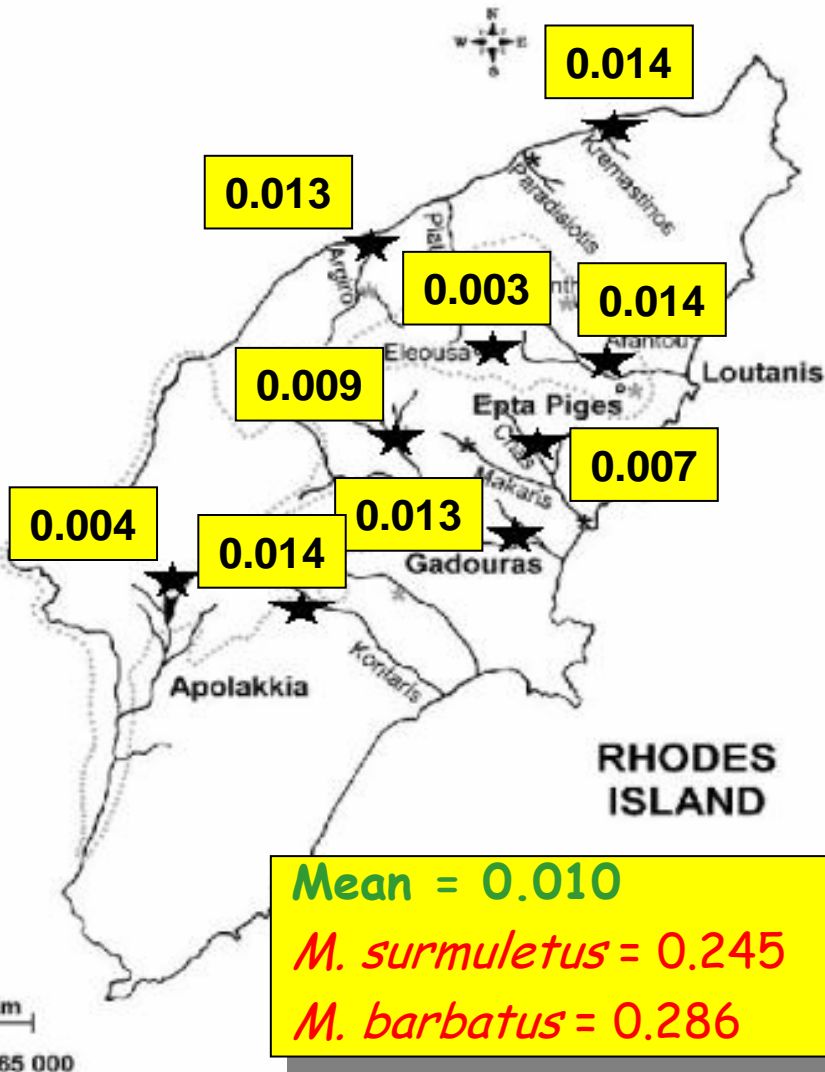


# A network indicating the relationships among the 6 haplotypes

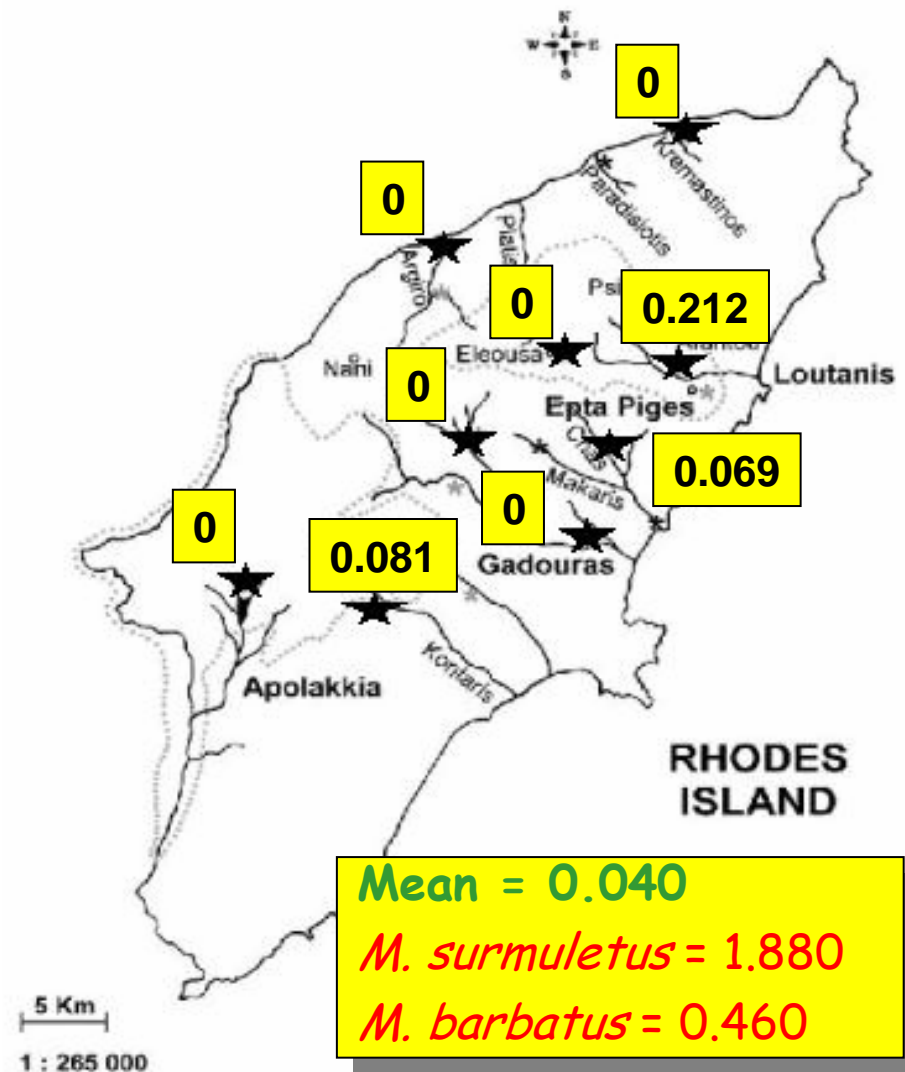


# A. Low genetic variation within populations

Low values of estimated heterozygosity for the RAPD



Low values (%) of nucleotide diversity for the mtDNA



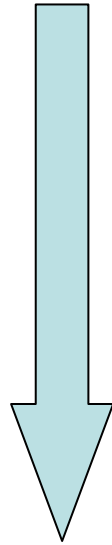


# A. Low genetic variation within populations

| Species                    | Mean Heterozygosity (RAPD)     | Mean Nucleotide Diversity (mtDNA)     |
|----------------------------|--------------------------------|---------------------------------------|
| <i>L. ghigii</i>           | 0.010<br>(this study)          | 0.040<br>(this study)                 |
| <i>Mullus surmuletus</i>   | 0.245<br>(Mamuris et al. 1999) | 1.880<br>(Mamuris et al. 2001)        |
| <i>Mullus barbatus</i>     | 0.286<br>(Mamuris et al. 1998) | 0.460<br>(Mamuris et al. 2001)        |
| <i>Silurus aristotelis</i> | -                              | 0.046<br>(Triantafyllidis et al 1999) |
| <i>Silurus glanis</i>      | -                              | 0.034<br>(Triantafyllidis et al 1999) |

Low levels of intra-population variability have been reported for other freshwater fish species in Greece: *Salmo trutta* (Apostolidis et al., 1996), *Leuciscus cephalus* (Imsiridou et al., 1998) and in Europe: *Nannoperca oxleyana* (Hughes et al., 1999), *Chondrostoma lusitanicum* (Mesquita et al., 2001).

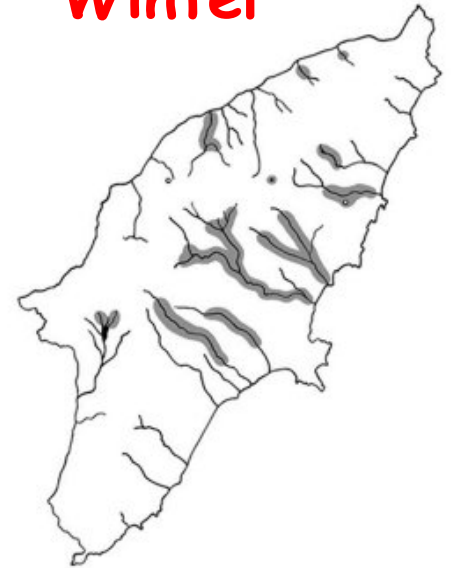
Successive bottleneck events evident in shrinkage and expansion of the populations year after year



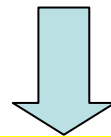
Summer



Winter



Complete loss of several genotypes and haplotypes, and an increased degree of inbreeding



Extremely low level of intra-population polymorphism revealed by both nuclear and mitochondrial DNA analysis

## B. Strong genetic structuring among populations

Nei's genetic distance based on RAPD (in blue) and nucleotide divergence ( $\times 10^2$ ) for mtDNA analysis (in green),

Lowest value      Highest value

| Populations    | 1     | 2     | 3     | 4     | 5     | 6     | 7     | 8     | 9     |
|----------------|-------|-------|-------|-------|-------|-------|-------|-------|-------|
| 1. Apolakkia   | -     | 0.410 | 0.509 | 0.825 | 0.410 | 0.410 | 0.410 | 0.424 | 0.420 |
| 2. Gadouras1   | 0.062 | -     | 0.095 | 0.410 | 0.000 | 0.000 | 0.000 | 0.013 | 0.007 |
| 3. Loutanis    | 0.092 | 0.029 | -     | 0.439 | 0.095 | 0.095 | 0.095 | 0.107 | 0.103 |
| 4. Eleoussa    | 0.083 | 0.051 | 0.044 | -     | 0.410 | 0.410 | 0.410 | 0.423 | 0.420 |
| 5. Argiros     | 0.053 | 0.036 | 0.046 | 0.054 | -     | 0.000 | 0.000 | 0.013 | 0.007 |
| 6. Gadouras2   | 0.067 | 0.002 | 0.029 | 0.053 | 0.033 | -     | 0.000 | 0.013 | 0.007 |
| 7. Kremastinos | 0.081 | 0.017 | 0.049 | 0.037 | 0.051 | 0.019 | -     | 0.013 | 0.007 |
| 8. Kontaris    | 0.046 | 0.031 | 0.050 | 0.064 | 0.044 | 0.030 | 0.050 | -     | 0.020 |
| 9. Chas        | 0.057 | 0.022 | 0.051 | 0.042 | 0.017 | 0.017 | 0.036 | 0.034 | -     |

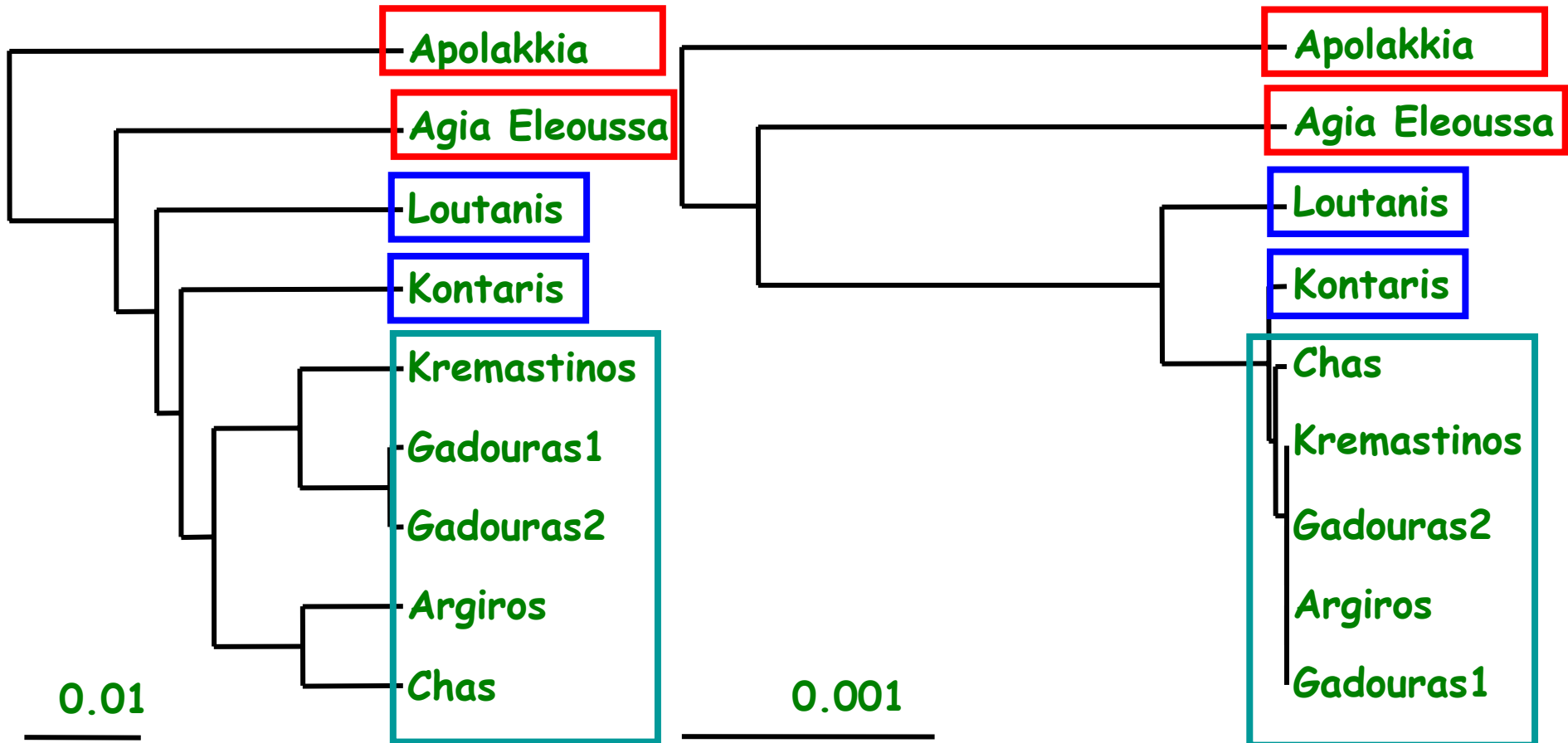
## B. Strong genetic structuring among populations

| Species                    | Mean Nei's genetic distance (RAPD) | Mean nucleotide divergence (%) (mtDNA)      |
|----------------------------|------------------------------------|---|
| <i>L. ghigii</i>           | 0.045<br>(this study)              | 0.206<br>(this study)                       |
| <i>Mullus surmuletus</i>   | 0.018<br>(Mamuris et al. 1999)     | 0.055<br>(Mamuris et al. 2001)              |
| <i>Mullus barbatus</i>     | 0.013<br>(Mamuris et al. 1998)     | 0.002<br>(Mamuris et al. 2001)              |
| <i>Silurus aristotelis</i> | -                                  | 0.000-0.110<br>(Triantafyllidis et al 1999) |
| <i>Silurus glanis</i>      | -                                  | 0.000-0.520<br>(Triantafyllidis et al 1999) |

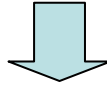
# UPGMA phenograms clustering the nine populations

Nei's (1978) genetic distance matrix, RAPD analysis

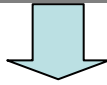
Net nucleotide divergence matrix mtDNA analysis



**It is probable that all haplotypes and/or alleles found here were initially represented in all populations**



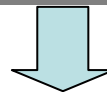
**Long time isolation coupled with successive bottleneck and subsequent genetic drift phenomena**



**Common mtDNA haplotypes and genotypes among the populations may have become rare or extinct through stochastic lineage loss**



**Because of the extreme physical isolation between certain sites and the absence of restocking operations**



**Bottleneck-generated genetic distance could not have been restored by subsequent genetic flow**



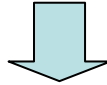
## **Suggestions for conservation**

**The appropriate unit of conservation for an endangered species may be evolutionarily significant units (ESUs)**

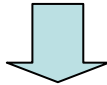
**Units on independent evolutionary trajectories justified by phylogeographic differentiation at the mtDNA level and significant divergence of allele frequencies at nuclear loci**

# Suggestions for conservation

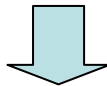
Fixation of RAPD markers and/or mtDNA haplotypes



Substantial genetic isolation of populations



The populations may be on independent evolutionary trajectories



Considering that each population appears well adapted so far within each site

**IT IS PROPOSED THAT ALL POPULATIONS SHOULD BE MANAGED AND CONSERVED SEPARATELY**

# Suggestions for conservation

Loss of genetic variability is a major threat to the survival of any species, decreasing the species' potential for adaptation to environmental changes

However

Both RAPD and mtDNA data showed that genetic variability within populations is already extremely low

Therefore

**TRANSLOCATIONS OF INDIVIDUALS, ATTEMPTING TO PREVENT A DRAMATIC DECLINE IN POPULATION DENSITIES, ARE NOT RECOMMENDED**

They could lead to transmission of pathogens, parasites and locally maladapted individuals or genes, without any certain benefit to increase of genetic variability

# Conservation actions concerned mainly:

✓ artificial breeding



A female and two males during fertilization in aquaria-indoor installations

✓ maintenance of fish stocks

constructions for the refuges in Loutanis stream



✓ planning of fish stockings